


Minne Heringa, October 29 2007

kiwa Partner for progress



Measurement of genotoxicity in (drinking) water

3^d NORMAN workshop


Monitoring of (drinking) water

- Pollution of ground water and surface water → drinking water quality?
- Current monitoring
 - Chemical monitoring
 - Biological monitoring
- New
 - Bio-assays: exposure of cells *in vitro*



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Which bioassays?

- Human health
- Low concentrations → no acute effects, sensitivity issue
- Life-time exposure → chronic toxicity most relevant

- Genotoxicity
- Endocrine disruption
- Other chronic toxicity?
 - Reproductive tox, carcinogenicity, neurological disorders, liver toxicity, kidney toxicity, etc.

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Which bioassays?


- Low concentrations → no acute effects, sensitivity issue
- Life-time exposure → chronic toxicity most relevant

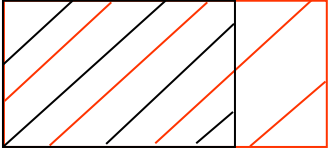
- **Genotoxicity**
- Endocrine disruption
- Other chronic toxicity?
 - Reproductive tox, carcinogenicity, neurological disorders, liver toxicity, kidney toxicity, etc.

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Genotoxicity and carcinogenicity

- **Genotoxic = causes DNA damage**
- **Carcinogenic = causes cancer**
 - Genotoxic carcinogens
 - Non-genotoxic carcinogens: e.g. growth promoters
- **Test for genotoxicity**
- **Test for non-genotoxic carcinogenicity?**





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Why test for genotoxicity?

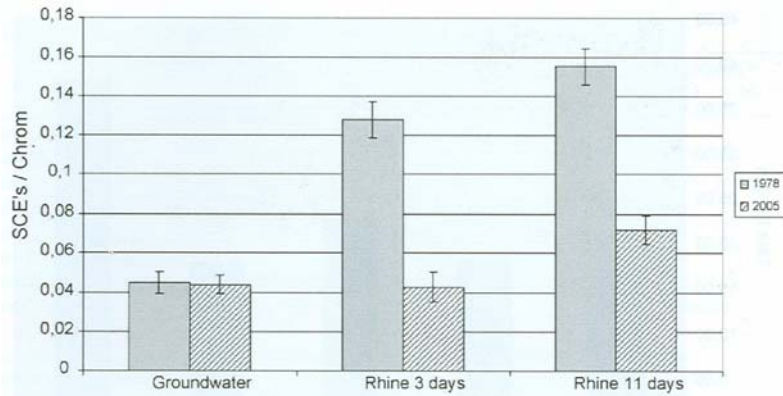
Compound	Log K _{ow}	Genotoxicity/ Carcinogenicity	Detected in*
Phenol	1.8	Genotoxic in mammalian cells	S
2,4-D	2.8	Genotoxic in mammalian cells	S
atrazine	2.5	Suspected genotoxicant	S, D
simazine	2.5	Genotoxic in mammalian cells	S, D
chlortoluron	2.3	Genotoxic in mammalian and bacterial cells	S
diuron	2.8	Genotoxic in mammalian and bacterial cells	S, D
trichloromethane	2.0	IARC 2B**	S, D
romodichloromethane	1.88	Suspected genotoxicant	S, D
ibromochloromethane	2.2	Suspected genotoxicant	S, D
tribromomethane	2.38	Suspected genotoxicant	S, D
tetrachloroethene	1.7	IARC 2B**	S, D
dimethylamine	-0.38	Genotoxic in mammalian cells	S
urotropine	-2.3	Genotoxic in mammalian cells	S
NDMA	-0.57	Carcinogenic (IARC 1)	S

Abstracted from van Genderen et al., *Inventory and toxicological evaluation of organic micropollutants*, report RIWA 2000

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Why test for genotoxicity?

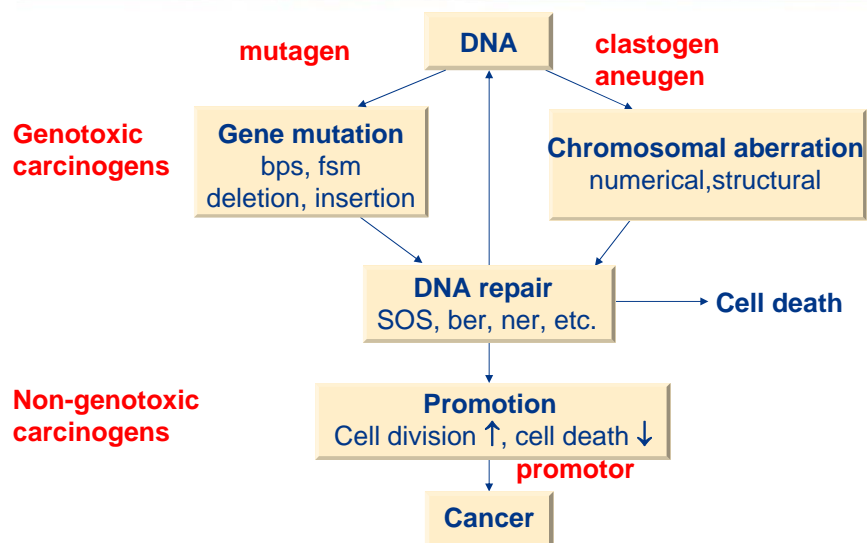
- Alink et al. *Mut Res* 631 (2007), p.93-100:
 - Eastern mudminnow exposed to Lekkanaal surface water
 - After 11 days significant genotoxic effects



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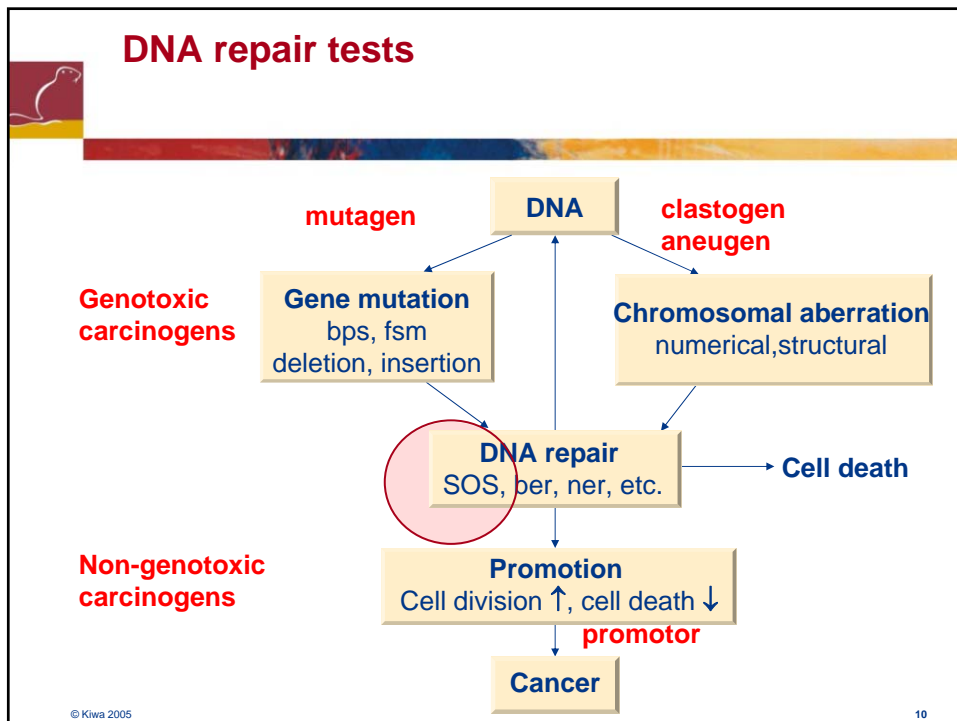
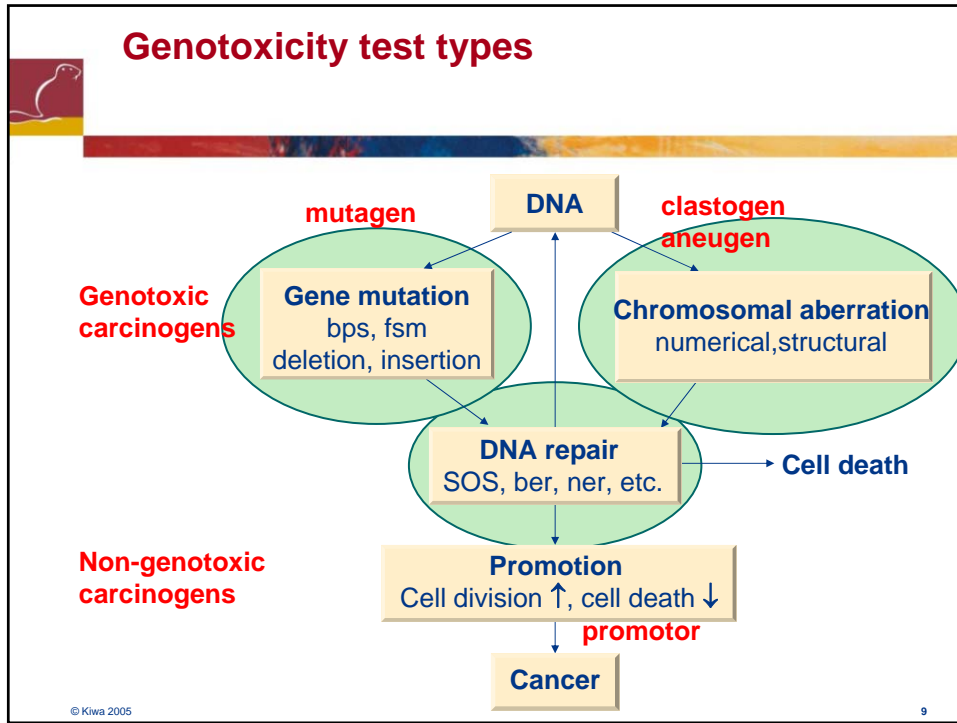
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Mechanism of carcinogenicity → types of genotoxicity



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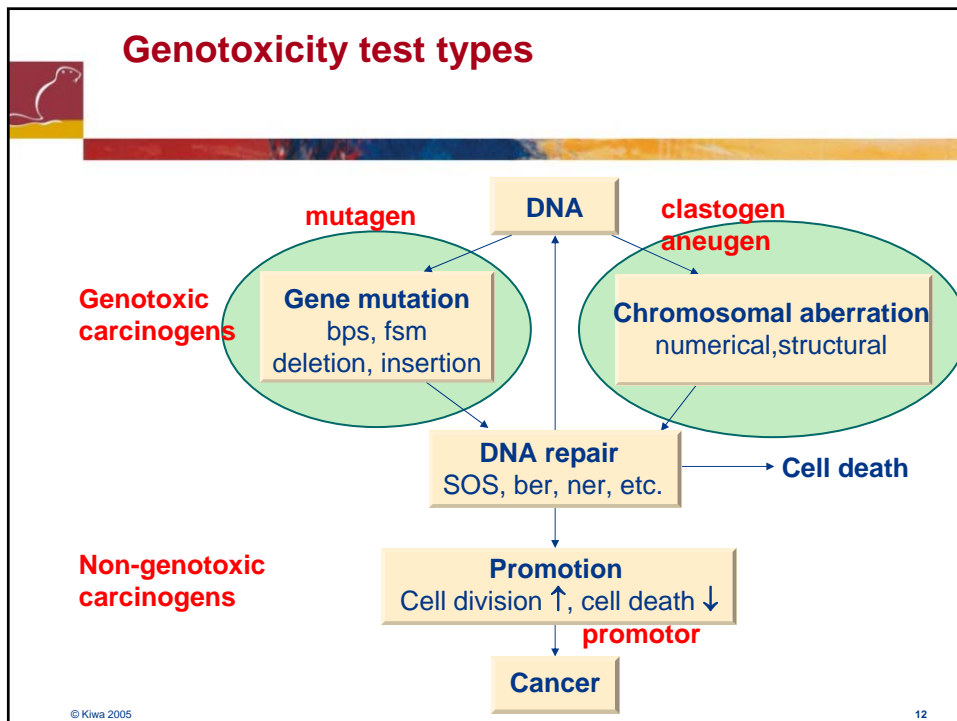
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Overview of DNA repair tests

- Well known repair assays
 - Unscheduled DNA synthesis (UDS)
 - Umu
 - SOS-Chromo
 - Vitotox®
 - Mutatox™
 - GreenScreen® (yeast and human)
 - RadarScreen

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Mutagenicity assays

- **Ames test / Ames II**

pathogenic bacteria, 3-4 days, colony counting

- ***Vibrio harveyi* neomycin resistance**

safe bacteria, 3-4 days, colony counting, not well known

- **HGPRT**

mammalian cell line, ± 2 weeks, colony counting

- **TK / mouse lymphoma**

special mammalian cell line, min. 2 weeks, colony counting



From www.hydrotox.de

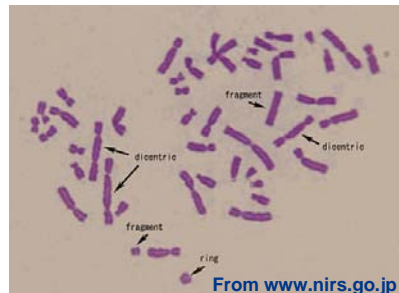
Chromosome aberration assays

- **Sister chromatid exchange (SCE)**

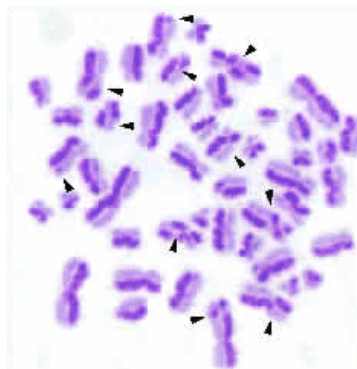
Colouring, microscope evaluation of every chromosome

- **Chromosome aberration (CA)**

Colouring, microscope evaluation of every chromosome



From www.nirs.go.jp



Sister Chromatid Exchange
Illustration produced in the laboratory of
Dr Al Rowland, Massey University

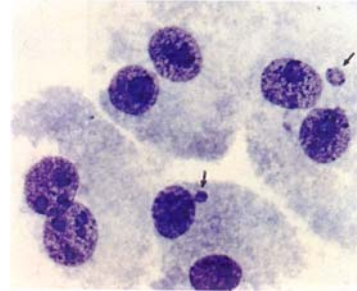
Chromosome aberration assays

■ Micronucleus

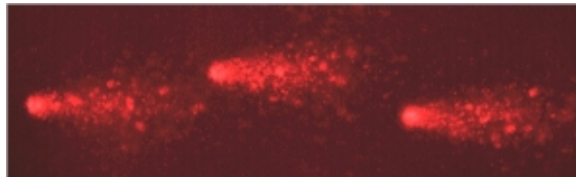
Cell division arrest, colouring,
microscope ev. of binucleated cells
(software)

■ Comet

Lysis, electrophoresis, microscope
ev. of cell (software)



Source: Darroudi et al., 1996



Strategy *in vitro* genotoxicity assays

■ Kiwa WR:

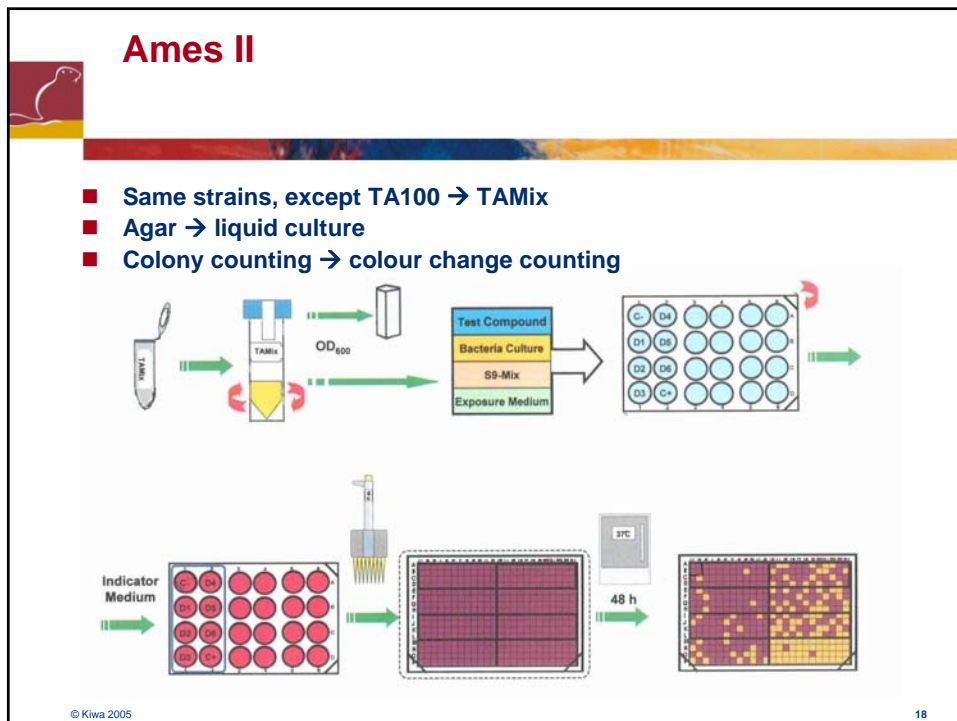
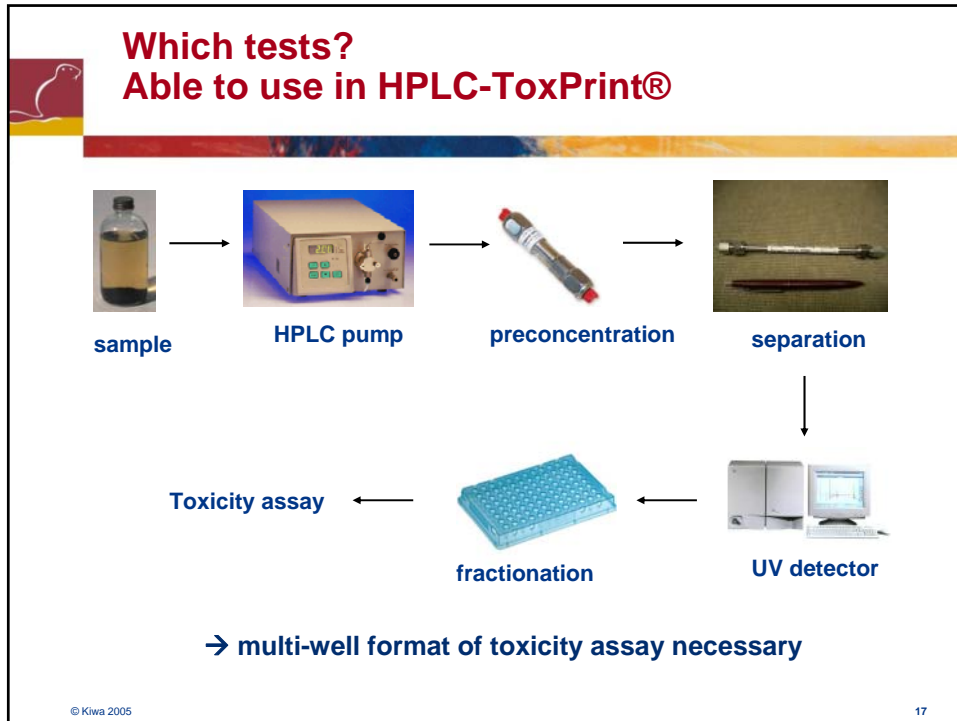
stage 1: bacterial mutagenicity + chrom. aberr.
stage 2: mammalian mutagenicity / other chrom. aberr

■ Gezondheidsraad (Health Council, NL; 1988):

bacterial mutagenicity + mammalian mutagenicity +
chromosomal aberrations

■ German working group GUM (2007):

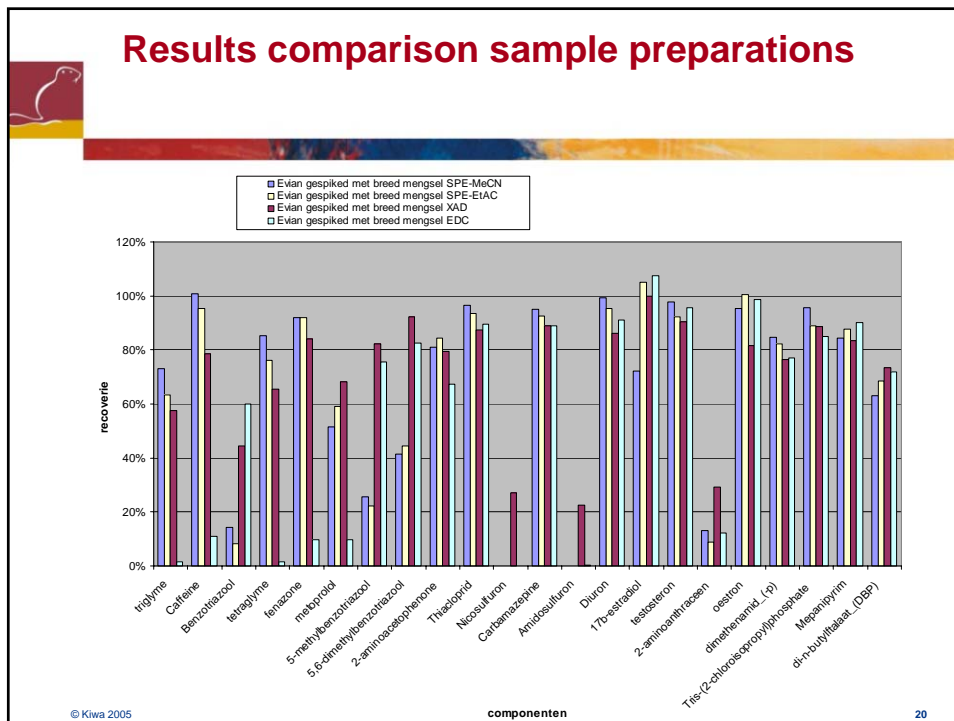
stage I: bacterial mutagenicity + *in vitro* micronucleus
stage II: *in vivo*



Adaptations Ames II for environmental samples: sample preparation

- **Extraction and concentration (10.000x) of water**
- **Compared different methods at ambient pH:**
 - **XAD SPE** classic, practical limitations
 - **SPE with OASIS HLB** new, easy
 - Elution with ACN general
 - Elution with EtAC hormones
 - **LLE with EtAC (for hormones)** classic, practical limitations
- **Water spiked with broad mixture**
- **Recoveries analyzed by LC-MS**

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Adaptations Ames II for environmental samples: statistics

- Low levels → when significant positive result?
- Statistical method

$$DL_{response} = \overline{NC} + t_r \cdot \left(\frac{s_r}{\sqrt{m}} \right)$$

$\overline{DL}_{response}$	= response detection limit
\overline{NC}	= average response negative control in test
t_r	= student t-value, from reproducibility test
s_r	= st. dev. of neg. control in reproducibility test
m	= number of replicates

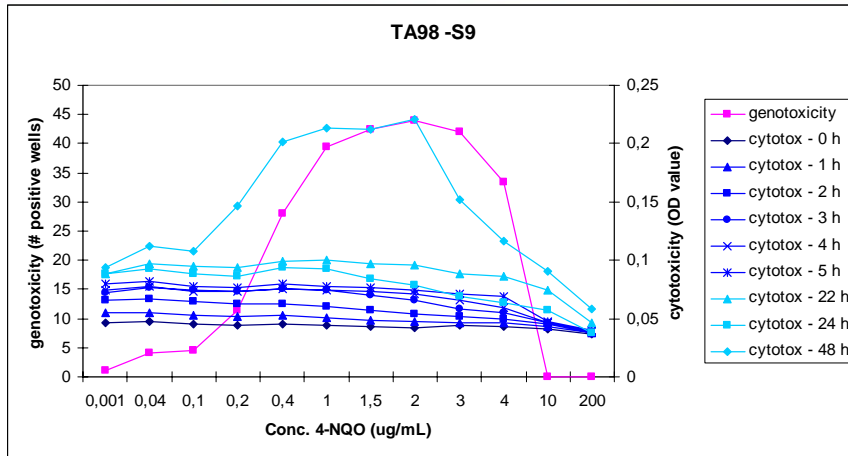
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Check for cytotoxicity

- No dose-response curve → check cytotoxicity
- Designed new method

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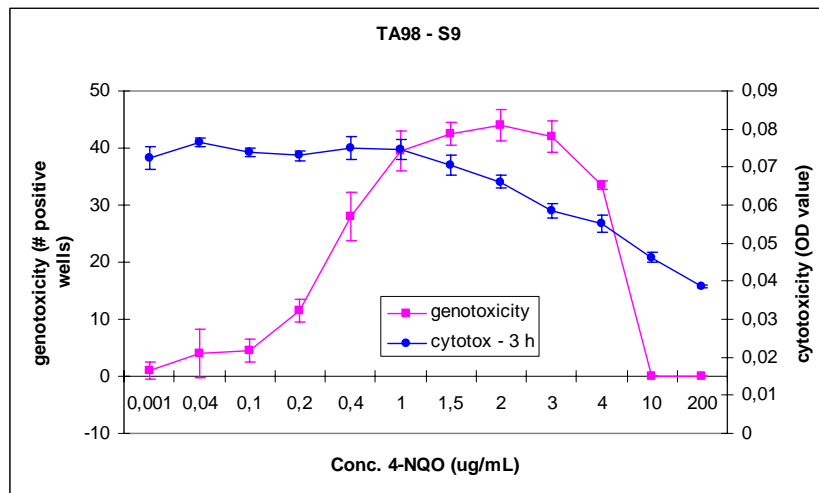
Optimal incubation time cytotoxicity method



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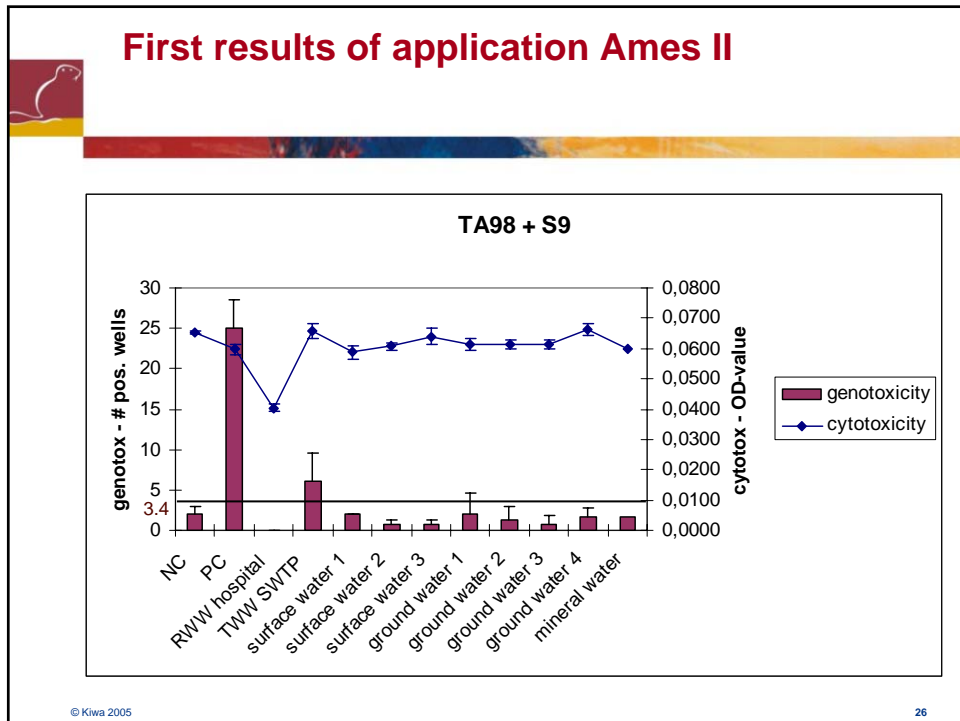
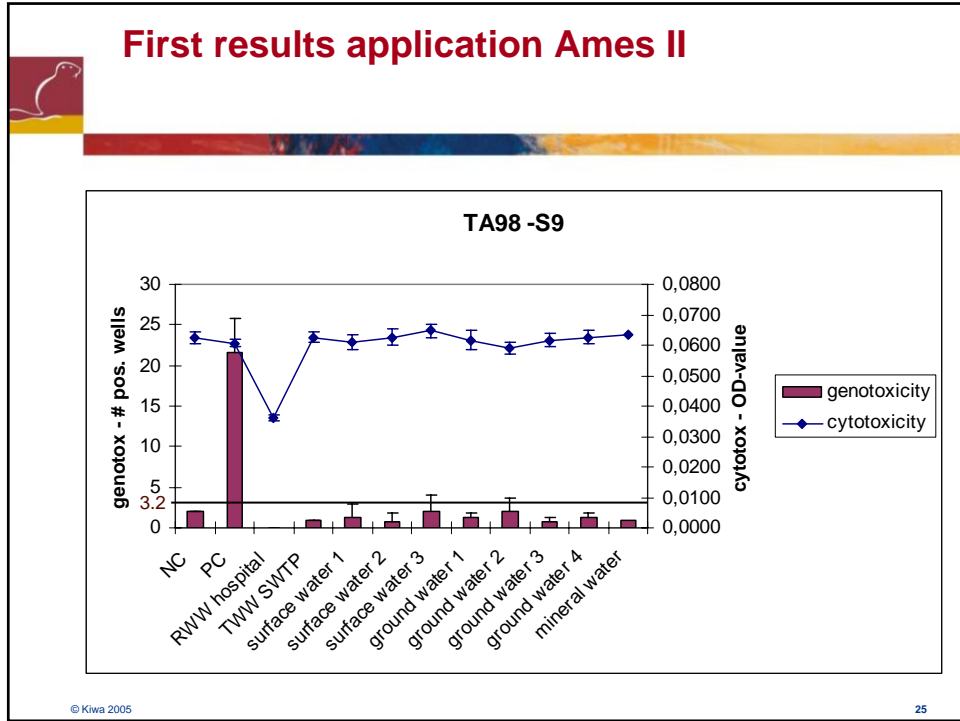
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Results cytotoxicity method



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Present and future research

- Choice between Comet and micronucleus
- What level of test response is unacceptable?
- Toxicity sensors with GMOs



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Toxicity sensor with GMOs

- Bacteria luminesce after exposure to toxicants
 - Plasmid with *promotor* - *luxCDABE*
- Panel of strains to cover all necessary toxic effects
 - Different promoters → different strains → respond to different toxicities
 - Also human toxicity
- Response in 1 hour → continuous monitoring
- Immobilization on optical fibres simplifies and eases regulatory acceptance
 - Developed by Ben-Gurion University from Israel
 - Kiwa WR has license to perform field test with these GMO-sensors

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Minne Heringa, October 29 2007



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